



Whole genome colony PCR

21st April 2021

This protocol describes a method to extract and prepare genomic and plasmid DNA from a bacterial colony. This has been verified using gram-negative *E. coli*. A colony was picked from a plate of cultured bacteria and then treated with Proteinase K before preparing libraries for sequencing using the Rapid PCR Barcoding Kit (SQK-RPB004). The sequencing performance of the prepared libraries was assessed using GridION.

Materials

- Plated bacterial colonies
- Rapid PCR Barcoding Kit (SQK-RPB004)
- SFB Expansion (EXP-SFB001)
- Flow Cell Priming Kit (EXP-FLP002)
- 1.5 ml Eppendorf DNA LoBind tubes
- 0.2 ml thin-walled PCR tubes
- Nuclease-free water (e.g. ThermoFisher, cat # AM9937)
- Agencourt AMPure XP beads
- [LongAmp Taq 2X Master Mix](#) (e.g. NEB M0287)
- 10 mM Tris-HCl pH 8.0
- 10 mM Tris-HCl pH 8.0, 50mM NaCl (or EB from Sequencing Auxillary Vials (EXP-AUX001 or equivalent)
- (Optional) [Thermolabile Proteinase K](#) (e.g. NEB P8111S)
- [Thermolabile Exonuclease I](#) (e.g. NEB M0568S)
- Microfuge
- Thermal cycler
- Pipettes and tips for P1000, P200, P100, P20, P10, and P2
- Timer
- Ice bucket with ice
- Magnetic separator, suitable for either 1.5 ml Eppendorf tubes or 0.2 ml PCR tubes
- Hula mixer (or similar gentle rotator mixer)
- DNA QC equipment, e.g. Agilent Bioanalyzer, Qubit fluorometer
- Inoculating loop/needle or sterile toothpick

Method

● Step 1

Using either a pipette tip, sterile toothpick, inoculating loop or needle, pick one colony and dip into 50 μ l of 10 mM Tris-HCl pH 8.0. Swirl for 10 seconds until the solution becomes turbid.

● Step 2 (enriching for plasmid DNA only)

If you are interested in the genomic DNA, proceed straight to step 4. If you are interested in the plasmid DNA, transfer the 50 μ l cell suspension to a 0.2 ml thin-walled PCR tube and incubate at 95°C for 5 minutes.

Note: It is observed that pre-treating the colony by heating leads to an enrichment in the observation of plasmid reads in the downstream sequencing, compared with non-heat treated libraries.

● Step 3 (enriching for plasmid DNA only)

Add 1 μ l of Thermolabile Proteinase K and incubate at 37°C for 15 minutes, then 55°C for 10 minutes.

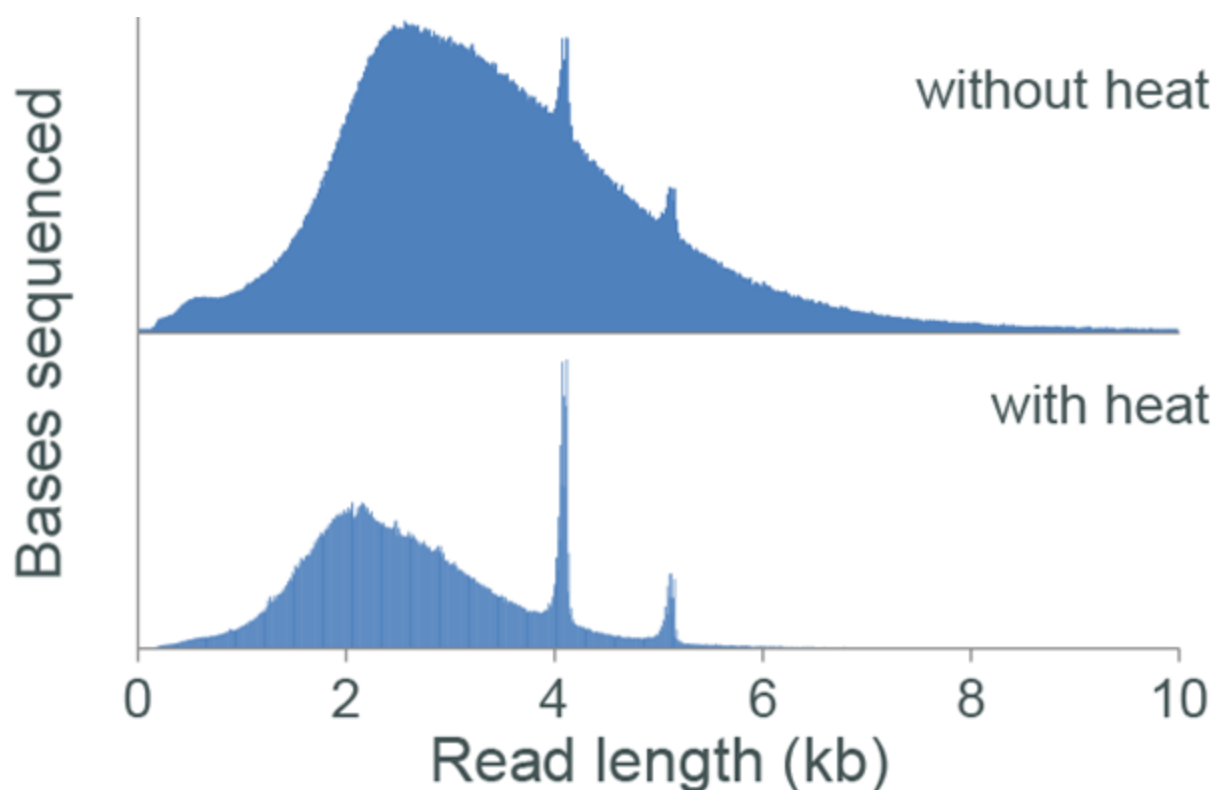
● Step 4

Libraries can be prepared for sequencing using the Rapid PCR Barcoding Kit (SQK-RPB004), using 3 μ l of the treated cell suspension as the template. If using cells without the heat treatment in step two, it is recommended to PCR for 25 cycles, and if using heat-treated cells, it is recommended to PCR for 30 cycles.

Sequencing performance

Libraries were prepared using the Rapid PCR Barcoding Kit (SQK-RPB004):

- Post PCR yield: 30-60 ng/μl
- Typical output: ★★ ★. Output from the flow cell may be increased by performing a [flow cell wash step](#) at the point where the rate of data acquisition begins to deteriorate due to the accumulation of pores in the “unavailable” or “recovering” state, and then adding a new library.
- Read length profile:



Date	Change note
21st April 2021	Title changed from 'PCR amplification of

	gram-negative bacterial DNA direct from a colony' to 'Whole genome colony PCR'. Updated step 3 sub-title from '(plasmid DNA only)' to '(enriching for plasmid DNA only)'.
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